DNA extraction from linseed containing samples using SureFood® PREP Basic

February 2025

1. Sample preparation

Please prepare the samples in duplicates

2. Additional required equipment and materials

- micro balance for weighing the sample
- 2.0 ml reaction tubes (not provided with the kit)
- heating block (up to 65°C)
- micro centrifuge (up to 12,000 rpm)

3. Procedure

- Transfer 50 mg sample material into a 2.0 ml reaction tube
- Add 800 μl Lysis Buffer and 40 μl Proteinase K
- Mix briefly on a Vortex mixer
- Incubate on a heating block under continuously shaking for 30 min at 65°C, according to the User Manual step <u>2 Lysis of the basic material</u>
- Centrifuge the sample lysate 1 min at 12,000rpm
- Place a Spin Filter (Code F, for pre-filtration) into a 2.0 ml Receiver Tube (Code R)
- Carefully transfer the supernatant direcly onto the Spin Filter, without the oil phase or precipitated proteins
- Centrifuge the Spin Filter with the Receiver Tube 1 min at 12,000rpm
- After centrifugation discard the Spin Filter
- Add 400 µl Binding Buffer to the filtrate and mix
- Place a Spin Filter (Code S, for binding) into a new 2.0 ml Receiver Tube (Code R)
- The binding of the nucleic acids on a Spin Filter takes place in two steps
 - 1. Transfer 650 μl of the filtrate-binding buffer mixture to the Spin Filter
 - 2. Incubate at room temperature for 1 min
 - 3. Centrifuge the Spin Filter with the Receiver Tube 1 min at 12,000 rpm
 - 4. Discard the filtrate and place the Spin Filter back into the Receiver Tube
 - 5. repeat the steps 1. to 4.
- Continue with step <u>5 Purification of the bound nucleic acids</u> from SureFood® PREP Basic